

# Role of in-utero Neem leaf (Azadirachta indica) supplemented diet on some hematological parameters and glucose storage in offspring of Wistar rats.

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Azadirachta indica (AI) Neem leaf is native of India which grows in most of tropical and subtropical countries. It tree has adaptability to a wide range of climatic, topographic and edaphic factors. This study explored the impact of AI supplementation during gestation on some haematological parameters and glucose storage in both offspring of Wistar rats. Eighteen pregnant female and 12 male Sprague-Dawley rats with a weight range of 140-180 grams were employed for this study and they were exposed to either a standard diet or AI supplementation (AIS). The pregnant rats were exposed to AIS up to birth (gestational AI supplementation) which comprised of both treated males (TM) and treated females (TF). Control rats with control diet was administered in analogous comparatively and this comprised of control male (CM) and control females (CF). During postnatal day 49, the rats were sacrificed and blood sample was obtained for assay of white blood cells (WBC), platelets (PLT), red blood cell counts (RBC), haemoglobin (Hb) and packed cell volume (PCV). Liver and gastrocnemius tissues were obtained for skeletal and hepatic glycogen assayed and intestinal and pancreatic  $\alpha$  amylase and  $\alpha$  glucosidase were determined from intestine and pancreatic tissues. WBC, PLT, RBC, Hb, PCV, alpha amylase, hepatic and skeletal glycogen increased significantly (p<0.05) in TM and TF with a remarkable decrease and increase in alpha glucosidase (p<0.05) in TM and TF respectively compared with CM and CF. It can be inferred from the present study that perinatal AI supplementation provides a substantial justification to its use in folk medicine as a hematopoietic plant and the increased glucose storage observed may not be unconnected with its role as an hypoglycaemic agent though the effect were more marked in female offspring compared with the male counterparts.

#### Introduction

Neem (Azadirachta indica), is native of India which is being cultivated in most of tropical and subtropical countries. It has adaptability to a wide range of climatic, topographic and edaphic factors. The tree survives well in dry, stony shallow soils and even on soils having hard calcareous or clay pan, at a shallow depth. So that it requires little water and plenty of sunlight to survive in the environment [1]. It is widely distributed throughout the world providing a source of inspiration for novel drug compounds, as plant derived medicines which have made large contributions to human health and well-being. Presently it can be seen growing successfully in over 70 countries worldwide, in Africa, Australia, Asia, North, Central and South America. Evidence showed that the tree was introduced into West Africa at the beginning of the present century [2]. Every part of the



tree has been employed as traditional medicine for household solution against various diseases [3]. It elaborates a vast array of biologically active compounds that are chemically diverse and structurally variable with different ingredients isolate from different parts of the tree [5]. The active ingredients include alkaloids, flavonoids, phenolic compounds, carotenoids, steroid and ketones, which have antihelminthic, antimicrobial, antiulcer, antifertility, antidiabetic, anti-inflammatory and antitumor properties and the plant is used in combination with oil for more effectiveness to reduce toxicity [5, 6,7]. The neem seed oil has toxicity effect against ectoparasites like ticks and mites which are common on cattle, equines, sheep, goats, wild ungulates and dogs [8]. Alcohol and aqueous extracts of flowers of the tree also effect against cattle filarial parasite [9].

All parts of the tree including leaves, bark, roots, seed and twigs contain active ingredients and used as medicine. Neem leaves are useful for increase immunity of the body, reduce fever, treating various foot fungi, useful against termites, used in curing neuromuscular pains and Anticlotting agent, Antihelminthic, Antituberculosis, antitumour, antiseptic, antiviral, Contraceptive, Cosmetics, Fertilizers, Insecticides and Insect repellents [12]. The processed Neem cake poses a good appetizer characteristic together with wormicidal activity which is used as poultry feed. Furthermore, Neem leaves has a significant amount of protein, minerals (except zinc) and digestible amounts of crude protein (CP) and total digestible proteins (TDP) which serves a better nutrition to the animals such as goat, sheep and cow [13]. Despite its bitter components, livestock consume diets containing varied percentage of neem cake. Alkali treatment of this by-product with caustic soda (10-20g sodium hydroxide) yields palatable products by removing the toxicant tritepenoids. After treatment it is incorporated into poultry feed [14].

Apart from their traditional uses, there are several reports on the biological activities and pharmacological actions of AI based on modern scientific investigations [15-21] blood glucose lowering activity of AI seed oil and leaf extracts have been reported in various models of diabetic animals [15-18] ethanol extracts of AI leaves have been shown to demonstrate antilipid peroxidative, antihyperglycemic and anti-hypercholesterolemic activities as well as reduced serum triglyceride level in diabetic rat model [18] Also, a significant decrease in some hematological parameters in chickens fed with AI leaves [19] and no significant difference in some hematological parameters following the extract use in diabetic rats [20] have been reported.

In Nigeria, AI supplements are popularly employed in the treatment of malaria. Some people have been observed consuming raw AI during pregnancy and lactation and anecdotal reports from them suggest that they consume it because of the myth that it is generally harmless, it is hematopoietic and protects them against malaria parasite. Despite these convincing evidences on its therapeutic use in folk medicine, however, there is paucity of information on the effects of AI supplemented diet during pregnancy and the subsequent effects on some hematological parameters and glucose storage in offspring. The present study therefore sought to unravel the role of AI supplementation on some hematological parameters and glucose storage and whether the effect is sex dependent.

#### Materials and methods

#### **Experimental animals**

Eighteen (18) pregnant Wistar rats with a weight range of 140-180g were employed and were sheltered with cages with quality lighting conditions 12hours light and dark cycles, and had free access to tap water with quality food and acclimatization lasted for 1week. The mechanism undertaken were in line with the presentations of the Experimentation Ethics Committee on Animals Use of the College of Medicine, University of Lagos, Lagos State in accordance with the United States National Academy of Sciences Guide for the Care and Use of Laboratory Animals.

#### Neem leaf (Azadirachta indica) AI collection



Fresh and matured AI leaves were collected from a tree within a community located in Afowowa in Ewekoro local government of Ogun State, Nigeria and was identified at the Biology unit of DS Adegbenro ICT polytechnic, Eruku-Itori, Ewekoro, Ogun state. The identified sample was deposited in the institution's herbarium. The collected samples were air dried and grinded with electric blender to get the powdered form.

#### Diet, mating and grouping

Five hundred grams of powdered AI leaves were grinded with 25kg of standard rat chow and this constitute AI supplemented diet with another 25kg normal rat chow and this formed the CONT diet. AI supplementation with these whole foods as a dried ground powder: maize (WN+M), sesame (WN+SO), soya bean meal (WN+SBM), palm kernel cake (WN+PKC), bone meal (WN+BM), industrial salt (WN+IS), grower premix (WN+GP), limestone (WN+LS), lysine (WN+L), methionine (WN+M), enzyme (WN+E), alphatox (WN+A) and threonine (WN+T). The diets gave a typical dietary intake of 1.5 servings of each food/d in a human diet, based on an energy content of a serving of food as set by the FDA for nutrition facts panels and a total daily diet intake of 2000kcal. Female rats were subjected to sexual act overnight with approved male breeders, 1 male per 2 females and they were maintained in their respective diet throughout pregnancy. The day on which spermatozoa was observed on a vaginal smear that was washed with normal saline NaCl 0.9% was assigned conception day 0. The rats were thereafter allocated to 1 of the 4 groups to be exposed to either a control diet or AI supplementation. Water and food were made available for all rats and grouped as follows (6 animals per group):

group I: Control male (CM) (exposed to control diet throughout the experiment).

group II: Control female (CF) (exposed to control diet throughout the experiment).

group III: Treated male (TM) (Male offspring exposed to AI supplementation only during gestation).

group IV: Treated male (TM) (Female offspring exposed to AI supplementation only during gestation).

All the pups were transferred to normal rat chow, except the control rats, until the end of the procedure which was PND 49. Offspring were reduced to 8-10 pups on postnatal PND 1 (birth, day 0) and were all weaned on PND 21 and housed in groups of 3 or 4, male and female offspring separately per cage.

#### Phytochemical analyses

The phytochemical screening of the plant was carried out on dried sample as described by [22] to identify the active components present in AI.

#### **Blood sample**

Four (4ml) of blood was obtained through cardiac puncture into an EDTA bottles and was centrifuged at 3000rpm for 15min with the plasma carefully obtained with rubber pippete in a clean Eppendorf bottle and were stored at -20 °C until analyses [23].

#### **Isolation of tissue**

On day 49 the rats were sacrificed via dislocation the of cervical vertebrate after a light anesthesia. The rats were dissected and skeletal, liver, pancreatic and intestinal tissues were obtained, washed in an ice cold and washed with 1.15 % KCl which was blotted and weighed [24].



#### **Hematological parameters**

Packed cell volume (PCV), hemoglobin Concentration (Hb), white blood cell count (WBC), red blood cell count (RBC) and platelet count (PLT); were analyzed using an Automated Analyzer (Sysmex, KX-21, Japan).

### Skeletal and hepatic glycogen

This was measured in both skeletal and liver tissue samples (homogenate) [23].

#### Assay of pancreatic and intestinal α-amylase

Tissue intestinal and pancreatic  $\alpha$ -amylase were assayed the method described by [24].

#### Assay of pancreatic and intestinal $\alpha$ -glucosidase

Tissue intestinal and pancreatic  $\alpha$ -glucosidase were assayed according to the method described by [24].

#### Statistical analysis

Results are provided as the mean and standard error of mean (SEM). GraphPad Prism 5 Software (GraphPad, Inc, La Jolla, CA, USA) was used for statistical analysis and one-way analysis of variance with post hoc Tukey's multiple comparison test was employed with significant level set at p<0.05.

#### **Results**

Table 1 showed the qualitative presence of phytochemicals which include tannins, Saponin, alkaloids, Flavonoids, Steroids, glycosides, Terpenoids and phenol.

Treated male (TM) and treated female (TF) investigated significantly increased (p<0.05) all blood parameters which include WBC (figure 1), PLT (figure 2), PCV (figure 3), RBC (figure 4), Hb (figure 5) assayed and also significantly decreased (p<0.05) for  $\alpha$ -amylase (figure 6) glycogen (figure 8) and hepatic glycogen (figure 9) contents compared with CM and CF.

SN	Phytochemicals	Result
1	Tannins	Positive
2	Saponin	Positive
3	Alkaloids	Positive
4	Flavonoids	Positive
5	Steroids	Positive
6	Glycosides	Positive
7	Terpenoids	Positive
8	Phenol	Positive

Table 1. Qualitative Phytochemicals Analysis of Powdered Neem Leaves Minerals Values 1.382mg/100g Tannins Saponin 2.914mg/100g Alkaloids 1.282mg/100g 4 1.073mg/100g Flavonoids 5 1.014mg/100g Steroids 6 2.134mg/100g Glycosides 4.356mg/100g Terpenoids



	P1 1	4 000 400
18	IPhenol	1.002mg/100g
8	1 1101101	1.0021119/1009

Table 2. Quantitative Phytochemicals Analysis of Powdered Neem Leaves

Table 2 revealed the presence of quantitative phytochemicals which include tannins (1.382mg/100g), Saponin (2.914mg/100g), alkaloids (1.282mg/100g), Flavonoids (1.073mg/100g), Steroids (1.014mg/100g), glycosides (2.134mg/100g), Terpenoids (4.356mg/100g), phenol (1.002mg/100g).

#### **Discussion**

The offspring treated with AI supplemented diet at in-utero showed an increase in the WBC, PLT, RBC, Hb and PCV. This observation is in agreement with the report of [25]. Also [26], a report from HIV/ AIDS patients, also observed that an acetone/water leaf extract of AI (IRAB) exhibited a remarkable increase in some hematological parameters. Pregnancy is a potentially anemic state [27,28] and since is often complicated by malaria in Nigeria and this might have effect on offspring health during postnatal life, this observed increase in hematological parameters in the present study suggests that the AI supplementation may have potentials at boosting erythropoiesis in offspring. Indeed, this may justify its use in folk medicine during pregnancy. The significance of the observed increased hematological parameters in the present study cannot be ruled out as anemia has been reported to affect over 500 million women [29] and in pregnancy it is associated with impaired maternal and infant consequences.

The heightened blood parameters could be related to the established constituents of the extract such as flavonoids and it is evident from the present study qualitative and quantitative presence of flavonoids that have been shown to have hematopoietic properties [28]. Also, AI has been established to boost the body's macrophage response, which stimulates the lymphatic system of the body and also boosts the body's leukopoiesis [30,31]

The effect of AI as a hypoglycemic agent in normal and experimentally induced diabetic animals is well established and reported [32-34]. The results of the present study also showed that the AI supplementation significantly increased the glucose storage in these rats which is evident form increased glucose deposition in both hepatic and skeletal tissues as well as increased activities of alpha amylase activity. This suggests that AI supplementation in all the treated rats, also possess hypoglycemic effect. Several mechanisms through which AI decreases the blood glucose level have been investigated by several authors [35,]. Jelodar, et al. [37], has earlier suggested that the hypoglycemic properties of the AI extract may not be unconnected with its ability to stimulate sufficient production of insulin by the pancreas, that aided in the peripheral utilization of glucose in the cells or the possible ability to regenerate the  $\beta$ -cells. Chattopadhyay, et al. [38], on the other hand suggested that AI's possible mechanism is by inhibiting the action of epinephrine on glucose metabolism resulting in increased utilization of peripheral glucose.

Alpha glucosidase is found in the mucosal brush border of small intestine where it catalyzes the final step of the digestion of starch and disaccharides which are present in abundance in human diet. Inhibitors of these enzymes delay the breakdown of carbohydrate in the small intestine and decreased the postprandial blood glucose movement levels in diabetic patients. Hence, these enzymes may be useful as effectiveness strategies to reduce the levels of post meal hyperglycemia [39].

#### **Conclusion**

In conclusion, the data described in the present study seem to provide a substantial justification the folkloric use of AI as a hematopoietic agent with the potential of increasing glucose storage in offspring though the effects re more marked in female offspring their male counterparts.



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#### References

- 1. Anonymous. Neem- Growing neem, organic farming, health, animal health, environmental use, home uses, economic potential, patents, new *e JBiio Electronic*; 2006.
- 2.Fathima SK. Investigations on the biology and management of Phomopsisazadirachtaeonneem [Ph.D. thesis]. Mysore, India: University of Mysore; 2004.
- 3. Hegde NG. Neem and small farmers constraints at grass root level. Indian JOR. 1995; 121:1040-8.
- 4. Kausik B, Ishita C, Ranajit K, Uday B. Biological activities and medicinal properties of neem (*Azadirachta indica*). Curr Sci. 2002;82(11):1336-45.
- 5. Subapriya R, Nagini S. Medicinal properties of neem leaves: a review. CMCACA. 2005;5(2):149-56. doi: 10.2174/1568011053174828.
- 6. Verkerk RHJ, Wright DJ. Biological activity of neem seed kernel extracts and synthetic *azadiracht* against larvae of Plutella xylostella. Pestic Sci. 1993;37(1):83-91. doi: 10.1002/ps.2780370113.
- 7. Parotta JA. Healing plants of Peninsular. India, New York: CABI Publishing. p. 495-96.
- 8. Choudhury M. Efficacy of neem leaf (Azadirachta indica, family: Meliaceae) in the control of larvae of Boophilusdecoloratus a onehosttick in cattle. J Nepal Pharm Assoc. -3.
- 9.Mishra V, Parveen N, Singhal KC, Khan NU. Antifilarial activity of *Azadirachta indica* on cattle filarial parasite Setaria cervi. Fitoterapia. 2005;76(1):54-61. doi: 10.1016/j.fitote.2004.10.010. PMID 15664463'
- 10.Gajalakshmi S, Abbasi SA. Neem leaves as a source of fertilizer-cum-pesticide vermicompost. Bioresour Technol. 2004;92(3):291-6. doi: 10.1016/j.biortech.2003.09.012, PMID 14766163.
- 11. Vethanayagam S, Rajendran S. Bioefficacy of neem insecticidal soap (NIS) on the disease incidence of bhendi, *Abelmoschus esculentus* (L.) Moench under field conditions. J of Biopest. 2010;3(1):246-9.
- 12.Prakash AO, Tewari RK, Mathur R. Non-hormonal post-coital contraceptive action of Neem oil in rats. J Ethnopharmacol. 1988;23(1):53-9. doi: 10.1016/0378-8741(88)90114-6, PMID 3419204.
- 13. Girish K, Neem SBS, Green Treasure A. J Biol. 2008;3:102-11.
- 14.Bawa G, Orumuyi A, Agbaji Z, Ladan, Okekeifi U. Effects of different methods of processing neem seed on performance of young growing rabbits. Pak J Nutr. -6.
- 15.Dixit VP, Sinha R, Tank R. Effect of neem seed oil on the blood glucose concentration of normal and alloxan diabetic rats. J Ethnopharmacol. 1986;17(1):95-8. doi: 10.1016/0378-8741(86)90076-0.
- 16.Halim EM. Lowering of blood sugar by water extract of Azadirachta indica and Abroma augusta in diabetes rats. Indian J Exp Biol. 2003;41(6):636-40. PMID <u>15266913</u>.



- 17. Gupta S, Kataria M, Gupta PK, Murganandan S, Yashroy RC. Protective role of extracts of neem seeds in diabetes caused by streptozotocin in rats. J Ethnopharmacol. 2004;90(2-3):185-9. doi: 10.1016/j.jep.2003.09.024, PMID 15013179.
- 18.Khosla P, Bhanwra S, Singh J, Seth S, Srivastava RK. A study of hypoglycaemic effects of Azadirachta indica (neem) in normal and alloxan diabetic rabbits. Indian J Phys Pharmacol. -74.
- 19.Ekaidem IS, Akpan HD, Usoh IF, Etim OE, Ebong PE. Effects of ethanolic extract of Azadirachta indica leaves on lipid peroxidation and serum lipids of diabetic Wistar rats. Acta Biol Szegedensis. -20.
- 20.Biu AA, Yusufu SD, Rabo JJ. Acute toxicity study on neem (*Azadirachta indica*, Juss) leaf aqueous extract in chicken (Gallus gallus domesticus). Afr Sci. 2010;11.
- 21.Itemobong SE, Atanghwo IJ, Akapan HD, Usoh IU, Etim OE, Ebong PE. Effects of ethanol extract of Azadirachta indica on some immunological and haematological parameters of diabetic Wistar rats. Afr J Pharm Pharmacol. -8.
- 22. Harborne. Phytochemical Methods. A guide to modern technology of plant analysis. 2nd ed. New York: Chapman & Hall; 1973. p. 88-125.
- 23. Morakinyo AO, Iranloye BO, Ogunsola OA. Glucometabolic effects of single and repeated exposure to forced-swimming stressor in Sprague-Dawley rats. Endocr Regul. 2018;52(2):85-92. doi: 10.2478/enr-2018-0010, PMID 29715186.
- 24.Igbayilola YD, Morakinyo AO, Iranloye BO. Adverse outcomes of perinatal protein restriction on glucose in offspring of Sprague-Dawley rats. Sci Afr. -11.
- 25. World Medical Association, American Physiological Society. Guiding principles for research involving animals and human beings. Am J Physiol Regul Integr Comp Physiol. 2002;283(2281-3. doi: 10.1152/aipregu.00279.2002, PMID 12121837.
- 26.Parshad O, Singh P, Gardner M, Fletcher C, Rickards E, Choo-Kang E. Effect of aqueous neem (*Azadirachta indica*) extract on testosterone and other blood constituents in male rats. A pilot study. West Indian Med J. 1994;43(3):71-4. PMID <u>7817539</u>.
- 27.Mbah AU, Udeinya IJ, Shu EN, Chijioke CP, Nubila T, Udeinya F, et al. Fractionated neem leaf extract is safe and increases CD4+ cell levels in HIV/AIDS patients. Am J Ther. 2007;14(4):369-74. doi:  $\underline{10.1097/MJT.0b013e3180a72199}$ , PMID  $\underline{17667213}$ .
- 28.O'Farrill-Santoscoy F, O'Farrill-Cadena M, Fragoso-Morales LE. Evaluation of treatment of iron deficiency anemia in pregnancy. Ginecol Obstet Mex. 2013;81(7):377-81. PMID <u>23971384</u>.
- 29. Townsley DM. Hematologic complications of pregnancy. Semin Hematol. 2013;50(3):222-31. doi:  $\underline{10.1053/j}$ .seminhematol. 2013.06.004, PMID  $\underline{23953339}$ .
- 30.Hanieh S, Ha TT, Simpson JA, Casey GJ, Khuong NC, Thoang DD et al. The effect of intermittent antenatal iron supplementation on maternal and infant outcomes in rural Viet Nam: A cluster randomised trial. PLOS Med. 2013;10(61001470. doi: <a href="https://doi.org/10.1371/journal.pmed.1001470">10.1371/journal.pmed.1001470</a>, PMID 23853552.
- 31.Raja SB, Murali MR, Kumar NK, Devaraj SN. Isolation and partial characterisation of a novel lectin from Aegle marmelos fruit and its effect on adherence and invasion of Shigellae to HT29 cells'. PLOS ONE. 2011;6(1):e16231. doi: 10.1371/journal.pone.0016231, PMID 21283697.



- 32.Sen P. Medira a PK, Ray A. Effects of Azadirachta indica A Juss on some biochemical, immunological and viscera parameters in normal and stressed rats. Indian J Exp Biol. -5.
- 33.Ray A, Banerjee BD, Sen P. 'Modulation of humoral and cell-mediated immune responses by Azadirachta indica (Neem) in mice'. Indian J Exp Biol. 1996;34(7):698-701. PMID <u>8979510</u>.
- 34. Shailey S, Basir SF. Strengthening of antioxidant defense by Azadirachta indica in alloxandiabetic rat tissues. J Ayurveda Integr Med. 2012;3(3):130-5. doi: 10.4103/0975-9476.100174, PMID 23125509.
- 35.Atangwho IJ, Ebong PE, Eyong EU, Asmawi MZ, Ahmad M. Synergistic antidiabetic activity of Vernonia amygdalina and Azadirachta indica: biochemical effects and possible mechanism. J Ethnopharmacol. 2012;141(3):878-87. doi: 10.1016/j.jep.2012.03.041, PMID 22521731.
- 36.Akhtar N, Khan BA, Majid A, Khan HM, Mahmood T, Gulfishan Pharmaceutical and biopharmaceutical evaluation of extracts from different plant parts of indigenous origin for their hypoglycemic responses in rabbits. Acta Pol Pharm. 2011;68(6):919-25. PMID 22125958'
- 37. Jelodar GA, Maleki M, Motadayen MH, Sirus S. Effect of fenugreek, onion and garlic on blood glucose and histopathology of pancreas of alloxan-induced diabetic rats. Indian J Med Sci. 2005;59(2):64-9. doi: 10.4103/0019-5359.13905.
- 38. Nandy AK, Podder G, Maitra SK. Preliminary report on antihyperglycemic effect of a fraction of fresh leaves of Azadirachta indica (Beng. Neem). Bull Calcua School. Trop Med. 1987; 35:29-35.
- 39. Teimori M, Montasser KS, Ghafarzadegan R, Hajiaghaee R. Study of Hypoglycemic Outcome of *Juglans regia* leaves and its Mechanism. J Med Plants. 2010; 9:57-65.